# ISOLATION AND CHARACTERIZATION OF A NEW GLABRETAL TRITERPENE FROM QUASSIA MULTIFLORA 

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#### Abstract

A new glabretal-type triterpene [1], was isolated from the roots of Quassia multiflora. Its structure was established by a combination of 2 D nmr experiments.


Quassia multiflora (A. Juss.) Nooteboom (=Simaba multiflora) is a member of the Simaroubaceae and has been widely investigated due mainly to its content of biologically active quassinoids (1-9). We have previously reported the isolation of quassinoids and squalene triterpenes from this plant $(5,10)$. We have investigated further extracts of Q. multiflora, and report here the isolation and characterization of a new glabretal-type triterpene whose structure has been established as 1. Glabretal triterpenes usually occur as epimeric mixtures at C-21 because most of them possess a hemiacetal in that position (11). However, since one epimer was present in a much higher concentration than the other, the characterization of $\mathbf{1}$ focused on the major component.

Compound $\mathbf{1}, \mathrm{C}_{35} \mathrm{H}_{56} \mathrm{O}_{7}$, isolated as a colorless gum, had ir absorptions characteristic of hydroxyl ( $3500 \mathrm{~cm}^{-1}$ ) and unsaturated ester ( $1703 \mathrm{~cm}^{-1}$ ) functionalities. The ${ }^{13} \mathrm{C}-\mathrm{nmr}$ spectrum displayed resonances for all 35 carbons of the major epimer. These included signals

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due to one hemiacetal at $\delta 97.53$ and five other $\mathrm{sp}^{3}$ carbons bearing oxygen at $\delta$ $78.72,78.14,74.99,74.22$, and 73.50. The hemiacetal carbon at $\delta 97.53$ was directly attached to a proton at $\delta 5.44(\mathrm{~d}$, $J=3.5 \mathrm{~Hz}$ ) as determined by an HMQC experiment. In the ${ }^{1} \mathrm{H}-\mathrm{nmr}$ spectrum, the protons of four oxymethines had signals at $\delta 4.68\left(W_{1 / 2}=7.5 \mathrm{~Hz}\right), \delta 4.48$ (dd, $J=8.5$ and 7.5 Hz$), \delta 3.76\left(W_{1 / 2}=8.0\right.$ $\mathrm{Hz})$, and $\delta 3.15(\mathrm{~d}, J=7.1 \mathrm{~Hz})$. Further, there were six tertiary methyls, in addition to a cyclopropyl methylene group with signals at $\delta 0.68$ and $\delta 0.48(2 \mathrm{H}$, $\mathrm{ABq}, J=4.3 \mathrm{~Hz}$ ). The presence of a tiglate group was evident from proton resonances at $\delta 6.86, \delta 1.86$, and $\delta 1.80$. The foregoing evidence suggested that compound 1 was a glabretal-type triterpene containing a tiglate ester.

A series of 2 D nmr experiments, which included DQF-COSY, HMQC, and HMBC sequences, led to the assign-
ment of all resonances for 1. In the HMBC experiment, methyl groups with resonances at $\delta 27.96(\mathrm{C}-28)$ and $\delta 21.82$ (C29) showed long-range correlations with each other, establishing their geminal disposition and that they were attached to C-4. In addition, these methyls showed long-range correlations to the oxymethine proton at $\delta 4.68$. Also, the tiglate carbonyl at $\delta 167.66$ displayed three-bond correlations with the same oxymethine at $\delta$ 4.68. These results indicated that the tiglate was attached to the oxygen at $\mathrm{C}-3$. The oxymethine signal at $\delta 3.76$ was located at $\mathrm{C}-7$ on the basis of the DQFCOSY and the HMBC experiments. These
experiments, along with the HMQC sequence, also revealed that the oxymethine carbon at $\delta 74.99$ was directly bonded to the proton at $\delta 3.15$ and that it was located at C-24. The stereochemistry at all relevant positions except C-24 was determined on the basis of analysis of a NOESY experiment, the hydroxyl group at $\mathrm{C}-21$ for the major epimer being $\beta$ oriented. These experiments led to the establishment of structure $\mathbf{1}$ for this new triterpenoid and the ${ }^{1} \mathrm{H}$ - and ${ }^{13} \mathrm{C}-\mathrm{nmr}$ assignments for this compound are reported in Table 1.

Compound $\mathbf{1}$ is closely related to a glabretal triterpene that was recently iso-

Table 1. Nmr Characteristics of Compound $\mathbf{1}$ in $\mathrm{CDCl}_{3}$ at 500 MHz .

| Position | $\delta_{C}$ | $\delta_{\mathrm{H}}\left(J_{\mathrm{HH}}\right)$ | HMBC |
| :---: | :---: | :---: | :---: |
| 1 | 34.17 | 1.38, 1.15 | 4.68, 0.90 |
| 2 | 26.28 | 1.92, 1.62 |  |
| 3 | 78.14 | 4.68 ( $W_{1: 2} 7.5$ ) | 0.90, 0.86 |
| 4 | 36.55 | - | 4.68, 2.05, 0.90, 0.86 |
| 5 | 41.26 | 2.05 | $4.68,3.76,0.90,0.86$ |
| 6 | 24.21 | 1.62, 1.58 | 3.76, 2.05 |
| 7 | 74.22 | $3.76\left(W_{12} 8.0\right)$ | 2.05, 1.62, 1.04 |
| 8 | 37.06 | - | 3.76 (w), 1.62, 1.04 |
| 9 | 44.20 | 1.30 | 3.76, 2.05, 1.04, 0.90 |
| 10 | 37.25 | - | 2.05, 1.62, 0.90 |
| 11 | 16.13 | 1.29, 1.29 |  |
| 12 | 22.72 | 1.91, 1.55 | 1.29 |
| 13 | 28.91 | - | $2.17,1.68,1.55,0.48$ |
| 14 | 38.91 | - | 1.04 |
| 15 | 25.71 | 2.08, 1.72 | 2.17, $0.68,0.48$ |
| 16 | 27.47 | 1.68, 1.68 | 2.17 |
| 17 | 44.96 | 2.17 | 1.97, 1.55, 0.48 |
| 18 | 13.74 | 0.68, 0.48 ( $\mathrm{ABq}, 4.3$ ) | 2.17, 1.91, 1.55 |
| 19 | 15.73 | 0.90 | 2.05, 1.30, 1.15 |
| 20 | 48.76 | 1.85 | 5.34, 4.48, 2.17, 1.97, 1.84 |
| 21 | 97.53 | 5.34 (3.5) | 4.48, 2.17, 1.85 |
| 22 | 29.48 | 1.97, 1.84 | 5.34, 4.48, 3.15, 1.85 |
| 23 | 78.72 | 4.48 (8.5,7.5) | $5.34,3.15,1.97$ |
| 24 | 74.99 | 3.15 (7.1) | $4.48,1.97,1.28,1.26$ |
| 25 | 73.50 | - | 1.28, 1.26 |
| 26 | 26.74 | 1.26 | 3.15, 1.28 |
| 27 | 26.73 | 1.28 | 3.15, 1.26 |
| 28 | 27.86 | 0.86 | 4.68, 0.90 |
| 29 | 21.82 | 0.90 | $4.68,2.05,0.86$ |
| 30 | 19.56 | 1.04 | 3.76, 1.30 |
| $1^{\prime}$ | 167.66 | - | 6.86, 4.68, 1.86, 1.80 |
| 2 '. | 129.26 | - | $6.86,1.86,1.80$ |
| $3^{\prime}$. | 136.62 | 6.86 (7.0, 1.5) | 1.86, 1.80 |
| $4^{\prime}$ | 12.20 | 1.86 (<1.0) | 6.86 |
| $5^{\prime}$. | 14.42 | 1.80 (7.0, <1.0) | 6.86 |

lated from Aglaia ferruginaea (Meliaceae) by Mulholland and Monkhe (11). The biosynthesis of glabretal triterpenes should involve a tirucallane precursor since all known examples to date have been isolated from plant families of the order Rutales (11-14), of which the tirucallanes are characteristic (15).

## EXPERIMENTAL

General experimental procedures.-It spectra were obtained on a Nicolet 3DX Ft-ir spectrometer. A Perkin-Elmer 243B polarimeter was used to obtain [ $\alpha]$ D values. Nmr spectra were recorded on a Varian Unity 500 spectrometer with TMS as internal standard. A VG-70-250S mass spectrometer was used to obtain ms data.

Plant material.-The plant material was collected at the Goethe Creek, Essequibo, Guyana, in November 1987. Voucher specimens (CAP 334) were deposited at the Jenman's Herbarium, University of Guyana.

Extraction and isolation.--The dried, ground roots ( 6.54 kg ) were extracted with $95 \%$ EtOH ( 93.5 liters). The extract was concentrated to a small volume ( 500 ml ), defatted with hexane ( $5 \times 200 \mathrm{ml}$ ), and subsequently extracted with $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ to yield a brown viscous syrup ( 167 g ) on removal of the solvent. This material, in portions, was chromatographed on Si gel using $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ with increasing amounts of MeOH as the solvent system to yield eight major fractions. Fraction 4 was rechromatographed on Si gel using hexane- $\mathrm{Me}_{2} \mathrm{CO}$ ( $3: 1$ ) as mobile phase to give $\mathbf{1}$ ( 32 mg ).

Compound 1.-Colorless gum; [ $\alpha$ ]D $-17.3^{\circ}$ ( $c=0.3, \mathrm{CHCl}_{3}$ ); ir $\nu \max 3500,1703 \mathrm{~cm}^{-1}$; eims m/z $570\left(\mathrm{M}^{+}-\mathrm{H}_{2} \mathrm{O}, 5\right), 552$ (3), 470 (12), 452 (11), 412 (23), 381 (18), 312 (28), 295 (25), 187 (58), 107 (44), 83 (100); hreims $m / z 570.3926$ calcd for $\mathrm{C}_{55} \mathrm{H}_{56} \mathrm{O}_{7}\left(\mathrm{M}^{+}-\mathrm{H}_{2} \mathrm{O}\right) 570.3920 ;{ }^{1} \mathrm{H}$ - and ${ }^{13} \mathrm{C}$-nmr data, see Table 1.

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